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June 5, 2019

Prince George's County Public Schools  
13300 Old Marlboro Pike  
Upper Marlboro, Maryland 20772  
Attention: Mr. Alex Baylor

RE: Indoor Air Quality Screening, Lake Arbor Elementary School  
IFB: 022-19  
ATI Project Number: ATI19-662

Dear Mr. Baylor:

Prince George's County Public Schools requested that ATI, Inc., conduct a proactive indoor air quality (IAQ) screening at Lake Arbor Elementary School. The IAQ screening was conducted on May 15, 2019. Its key findings are enclosed in the Executive Summary on page three, and the official laboratory report for total fungal spore trap sampling is enclosed in Appendix A.

Thank you for the opportunity to provide Industrial Hygiene services for Prince George's County Public Schools. If you have any questions regarding this report, please contact us at (202) 643-4283.

Sincerely,  
**ATI, INC.**

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Courtney E. McCall  
Project Manager

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Sarath Seneviratne  
CIH, CSP, CHMM

## Indoor Air Quality Screening Report



Prince George's County Public Schools  
Lake Arbor Elementary School  
10205 Lake Arbor Way  
Mitchellville, Maryland 20721

Prepared for:

Prince George's County Public Schools  
13300 Old Marlboro Pike  
Upper Marlboro, Maryland 20772

**June 5, 2018**

Submitted by:

**ati**

ATI Job # 19-662

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### Abbreviations and Acronyms

<b>AHU</b>	Air-Handling Unit
<b>AIHA</b>	American Industrial Hygiene Association
<b>ASHRAE</b>	American Society of Heating, Refrigerating and Air-Conditioning Engineers
<b>ASTM</b>	American Society for Testing and Materials
<b>CO</b>	Carbon Monoxide
<b>CO<sub>2</sub></b>	Carbon Dioxide
<b>EMLAP</b>	Environmental Microbiology Laboratory Accreditation Program
<b>HVAC</b>	Heating, Ventilating, And Air-Conditioning
<b>IAQ</b>	Indoor Air Quality
<b>NIST</b>	National Institute for Standards and Technology
<b>NVLAP</b>	National Voluntary Laboratory Accreditation Program
<b>RH</b>	Relative Humidity

Abbreviations involving scientific volume and measurements involving media or water sampling

<b>Counts/m<sup>3</sup></b>	Mold spores per cubic meter of air
<b>LPM</b>	Liters Per Minute
<b>NTE</b>	Not to exceed
<b>°F</b>	degree Fahrenheit
<b>PPM</b>	Parts Per Million

## 1. Executive Summary and Key Findings

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ATI conducted a proactive Indoor Air Quality (IAQ) screening on May 15, 2019, at Lake Arbor Elementary School, located at 10205 Lake Arbor Way, Mitchellville, MD 20721.

The screening included a visual assessment of randomly selected classrooms and other frequently occupied spaces, such as the cafeteria, the main office, and classrooms, for potential IAQ contributors and pathways. As part of the screening, ATI collected direct reading measurements for comfort parameters, including temperature, relative humidity, carbon dioxide, and carbon monoxide. Also, ATI collected total fungal air samples on spore trap cassettes for microbiological analysis.

The following is a summary of the key findings from this screening:

1. Temperature measurements were slightly below and on the lower end of the ASHRAE guidelines for summer temperatures, between 73°F and 79°F.
2. Humidity measurements were within ASHRAE guidelines, <65%.
3. Carbon dioxide measurements were within the ASHRAE maximum for the day of the screening, with one location exceeding 1,125 ppm.
4. Carbon monoxide was not detected throughout the tested spaces.
5. Laboratory analysis indicated that total fungal concentrations on the spore traps did not show significant indoor amplification. Indoor concentrations of mold compared favorably to those detected outdoors.

## 2. Assessment Methods

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Mr. Brian Chapman and Ms. Mikal Frater of ATI, Inc., conducted a visual assessment and air sampling on May 15, 2019. Sampled rooms were randomly selected and accounted for approximately 10% of classrooms, or a minimum of five samples. Visual observations were made at the time the samples were collected. ATI references the American Society of Heating, Refrigerating, and Air-Conditioning Engineers (ASHRAE) *Standard 62.1 – 2016* and *ASHRAE Standard 55 – 2017* when providing IAQ services to clients. ASHRAE is an industry leader on energy efficiency and indoor air quality.

All measurements and air samples were collected between three-six feet from floor elevation, which represents the breathing zone, and away from air supply and air return diffusers. Real-time direct readings for temperature, relative humidity, carbon dioxide (CO<sub>2</sub>), and carbon monoxide (CO), were obtained with a calibrated TSI Q-Trak 7575-X Meter and a 982 TSI probe.

Total fungal air samples were collected with a Buck BioAire High-Volume Sampling Pump on Zefon Air-O-Cell spore-trap cassettes at a flow rate of 15 liters per minute for five minutes, for a sample volume of 75 liters. The samples were analyzed by direct microscopic examination (identifies and counts both viable and non-viable spores, which is then considered “total fungal”), via the American Society for Testing and Materials (ASTM) Standard D7391-09 by EMSL Analytical, Inc., (EMSL) located in Beltsville, MD.

EMSL participates in the National Institute of Standards and Technology’s (NIST’s) National Voluntary Laboratory Accreditation Program (NVLAP) for general laboratory performance and management and the

American Industrial Hygiene Association (AIHA) Environmental Microbial Laboratory Accreditation Program (EMLAP, Certificate Number 102891).

Instrument calibration records are included in Appendix B of this report.

**3. Visual Observations**

**Table 1: Visual Observations and Sampling Locations**

Sample Location	Observations
Outside	<ul style="list-style-type: none"> <li>• Lawn mowing in distance.</li> <li>• Clear skies, light winds.</li> <li>• Light vehicle traffic in parking lot.</li> </ul>
Main Office	<ul style="list-style-type: none"> <li>• Medium foot traffic.</li> <li>• Four air diffusers. One air return.</li> <li>• 3-7 occupants in room during sampling.</li> <li>• Door to corridor open occasionally.</li> <li>• Office splits into 6-8 additional rooms.</li> <li>• Space is approximately 369 ft.<sup>2</sup></li> </ul>
Media Center	<ul style="list-style-type: none"> <li>• Two ceiling tiles with old water stains in reading nook near computer lab. One stain has valve above the plenum – stain has possible growth.</li> <li>• A lot of books are in this space.</li> <li>• Two occupants are in area during sampling.</li> <li>• Emergency exit in room – outside access.</li> </ul>
Room 203	<ul style="list-style-type: none"> <li>• One air return, one diffuser.</li> <li>• Wall unit supplies both A/C and heat.</li> <li>• Three occupants in area during sampling.</li> <li>• Light foot traffic.</li> <li>• Radon fan inside plenum.</li> <li>• Stained ceiling tile with growth.</li> <li>• Restroom inside classroom.</li> </ul>
Room 305	<ul style="list-style-type: none"> <li>• One air diffuser, one air return.</li> <li>• One wall unit.</li> <li>• Three occupants in room during sampling.</li> <li>• Space is approximately 960 ft.<sup>2</sup></li> </ul>
Room 334	<ul style="list-style-type: none"> <li>• Stale, stagnant air.</li> <li>• Humid space.</li> <li>• One wall unit. A/C not in operation.</li> <li>• Three occupants in room during sampling.</li> <li>• Door to corridor open mid-sampling.</li> <li>• Space is approximately 875 ft.<sup>2</sup></li> </ul>

Sample Location	Observations
Room 240	<ul style="list-style-type: none"> <li>• Smells of paint.</li> <li>• Emergency exit in room – outside access.</li> <li>• Moderate traffic.</li> <li>• One wall unit.</li> <li>• Twenty occupants in area during sampling.</li> <li>• Two air diffusers, one air return.</li> <li>• Space is approximately 1000 ft.<sup>2</sup></li> </ul>
Cafeteria	<ul style="list-style-type: none"> <li>• Four occupants in area during sampling.</li> <li>• Light traffic right after lunch.</li> <li>• Custodian sweeping cafeteria floor.</li> <li>• Outside access through cafeteria.</li> <li>• Door to corridor occasionally open.</li> <li>• Large occupied area.</li> </ul>

**4. Thermal Environmental Conditions for Human Occupancy**

ASHRAE Standard 55-2017, *Thermal Environmental Conditions for Human Occupancy*, addresses thermal comfort in an office environment, which means that an employee wearing a normal amount of clothing feels neither too cold nor too warm. This standard discusses thermal comfort within the context of air temperature, humidity, and air movement and provides recommended ranges for temperature and humidity that are intended to satisfy most building occupants. The recommended ASHRAE ranges are referenced below by each comfort parameter.

**4.1 Temperature**

The ASHRAE standard establishes a winter comfort range of between 68°F and 75°F and a summer range of between 73°F and 79°F. The temperature measurements obtained during the May 15, 2019 screening is summarized in Table 2. As indicated by the data in the table, temperatures in the school averaged between 71.5- 75.9°F, below and on the lower end of the ASHRAE summer comfort range.

**Table 2: Temperature Measurements**

Sample Location	May 15, 2019 °F			ASHRAE Standard °F
	Min	Max	Average	
Outside	72.0	75.0	73.5	N/A
<b>Indoors</b>				
Main Office	75.9	75.9	75.9	73 – 79
Media Center	70.6	72.4	71.5	73 – 79
Room 203	72.4	72.4	72.4	73 – 79
Room 305	72.2	72.2	72.2	73 – 79





Sample Location	May 15, 2019 °F			ASHRAE Standard °F
	Min	Max	Average	
Room 334	74.1	75.0	74.55	73 – 79
Room 240	72.5	72.5	72.5	73 – 79
Cafeteria	73.2	73.2	73.2	73 – 79

#### 4.2 Relative Humidity

Relative humidity is a key factor for mold growth. Mold has the potential of growing on suitable surfaces with humidity levels above 65%. ASHRAE *Standard 62.1-2016, Ventilation for Acceptable Indoor Air Quality*, recommends a maximum indoor relative humidity of 65% to prevent condensation of moisture on surfaces. Relative humidity below 30% may result in drying of the mucous membranes and skin. Relative humidity measurements are summarized in Table 3. As indicated by the data in the table, relative humidity measurements averaged between 36.4 and 52.95%, below the ASHRAE maximum recommendation of 65% relative humidity.

**Table 3: Relative Humidity Measurements**

Sample Location	May 15, 2019 (%)			ASHRAE Standard (% RH)
	Min	Max	Average	
Outside	24.4	25.0	24.7	N/A
<b>Inside</b>				
Main Office	36.2	36.6	36.4	< 65
Media Center	40.1	40.1	40.1	< 65
Room 203	38.6	38.6	38.6	< 65
Room 305	38.7	38.7	38.7	< 65
Room 334	51.8	54.1	52.95	< 65
Room 240	44.1	44.1	44.1	< 65
Cafeteria	44.2	44.6	44.4	< 65

#### 4.3 Carbon Dioxide

Carbon dioxide measurements within an occupied building are a standard method used to gauge the efficiency of ventilation systems. Carbon dioxide is a by-product of human respiration and does not pose an acute health hazard alone. Elevated concentrations may suggest that insufficient fresh air is being supplied to an occupied space and/or that the ventilation system does not provide a sufficient rate of air exchange.

Research has indicated that buildings with adequately operating ventilation systems are able to remove odors generated by activities in an indoor office environment efficiently. ASHRAE *Standard 62.1-2016* states that comfort (odor) criteria with respect to human bioeffluents are likely to be satisfied if the ventilation results

indoor carbon dioxide concentrations are less than 700 parts per million (ppm) above the outdoor air concentration.

Carbon dioxide measurements are summarized in Table 4. On the day of the screening, the average outdoor carbon dioxide concentration obtained was 425 ppm, which calculates to a maximum indoor concentration of 1,125 ppm (700 + 425). The carbon dioxide levels inside the suite ranged from the average minimum detected, 489 ppm to 2,640 ppm, the average maximum detected, with one location exceeding the maximum recommended concentration of 1,125 ppm.

**Table 4: Carbon Dioxide Measurements**

Sample Location	May 15, 2019 Concentration (parts per million)			ASHRAE Standard (ppm) NTE
	Min	Max	Average	
Outside	413	437	425	N/A
<b>Inside</b>				
Main Office	774	790	782	1,125
Media Center	682	686	684	1,125
Room 203	487	491	489	1,125
Room 305	520	524	522	1,125
Room 334	2,597	2,683	2,640	1,125
Room 240	946	1030	988	1,125
Cafeteria	952	970	961	1,125

**4.4 Carbon Monoxide**

Carbon monoxide is a colorless and odorless gas produced by the incomplete combustion of carbon containing fuels. Oil, gasoline, diesel fuels, wood, coke, and coal are the major sources of carbon monoxide. ASHRAE recommends that carbon monoxide not exceed nine ppm indoors. As indicated by the data in Table 5, carbon monoxide was not detected throughout the suite.

**Table 5: Carbon Monoxide Measurements**

Sample Location	May 15, 2019 Concentration (parts per million)			ASHRAE Standard (ppm)
	Min	Max	Average	
Outside	0	0	0	N/A
<b>Inside</b>				
Main Office	0	0	0	< 9
Media Center	0	0	0	< 9
Room 203	0	0	0	< 9
Room 305	0	0	0	< 9
Room 334	0	0	0	< 9



Sample Location	May 15, 2019 Concentration (parts per million)			ASHRAE Standard (ppm)
	Min	Max	Average	
Room 240	0	0	0	< 9
Cafeteria	0	0	0	< 9

## 5. Total Fungal Air Sampling Results

Mold needs a food source, moisture, proper temperature and humidity, and at most times, a source of light, to grow in an environment. Air filtration through building entrances and exits, open windows and loading docks, and foot traffic into buildings, serve as primary pathways that bring mold indoors. Water leaks and humid conditions inside of buildings provide the moisture that fosters mold growth.

The May 15, 2019, mold screening sampled air using spore trap cassettes in randomly selected classrooms and other rooms. These cassettes collect both viable spores, those capable of producing more fungal colonies, and non-viable spores, which cannot reproduce. Based upon recognized industry practices, indoor mold concentrations are compared with those detected outdoors, which are also known as ambient or baseline samples.

In normal circumstances, the diversity of spores identified indoors and outdoors should be similar with some exceptions. The dominating presence of one or two species of fungal spores identified indoors and the absence of the same species outdoors can indicate a moisture problem with the potential to degrade the air quality. Fungi species present indoors are typically found at levels ranging from approximately 10-50% of their levels in the outdoor air, reflecting the filtering by the building's HVAC system.

The official laboratory report with spore trap samples collected on May 15, 2019 is presented in Appendix A. The findings indicated that the indoor concentrations were favorable compared to the outdoor concentrations, and no tested space exceeded the total concentration detected outdoors, which was 8,370 counts/m<sup>3</sup>.

Ascospores, Basidiospores and Cladosporium were the predominant spore types detected indoors, yet they did not exceed the concentration found in the outdoor sample. These spores are commonly found in indoor environments and may cause allergies. Aspergillus/Penicillium, which can cause allergies and health problems for persons with more serious lung disorders, was also detected indoors but not at levels exceeding the outdoor sample. Ascospores, Basidiospores, Cladosporium and Aspergillus/Penicillium are not known to be water damage indicators.

## 6. Summary of Findings

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ASHRAE comfort parameters including relative humidity and carbon monoxide were within recommended ranges in all tested areas. The indoor temperatures fell below and on the lower end of the ASHRAE recommended summer comfort range. Carbon dioxide levels in one location exceeded the ASHRAE maximum for the day of the screening.

Indoor concentrations of mold compared favorably to those detected outdoors.

We appreciate the opportunity to provide these IAQ testing services for you. If you have any questions, please contact us at (202) 643-4283.

Best regards,  
**ATI, INC.**



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Courtney E. McCall  
Project Manager



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Sarath Seneviratne  
CIH, CSP, CHMM

**Appendix A:  
Laboratory Report and Chain of Custody**



# EMSL Analytical, Inc.

2500 Gateway Centre Blvd., Suite 600 Morrisville, NC 27560

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<http://www.EMSL.com> / [raleighlab@emsl.com](mailto:raleighlab@emsl.com)

<b>EMSL Order:</b> 291905207
<b>Customer ID:</b> ATII25A
<b>Customer PO:</b>
<b>Project ID:</b>

<b>Attn:</b> Brian Chapman ATI 4221 Forbes Blvd Suite 250 Lanham, MD 20706 <b>Project:</b> 19-662-PGCPs - Lake Arbor ES	<b>Phone:</b> (202) 368-1376 <b>Fax:</b> <b>Collected:</b> 05/15/2019 <b>Received:</b> 05/15/2019 <b>Analyzed:</b> 05/21/2019
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### Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number: Client Sample ID: Volume (L): Sample Location	291905207-0001 19-662-01 75 Outside Parking Lot			291905207-0002 19-662-02 Field Blank			291905207-0003 19-662-03 75 Main Office			
	Spore Types	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total
Alternaria (Ulocladium)	-	-	-	-	-	-	-	-	-	-
Ascospores	51	2200	26.3	-	-	-	1	40	3.2	
Aspergillus/Penicillium	23	970	11.6	-	-	-	2	80	6.3	
Basidiospores	70	3000	35.8	-	-	-	26	1100	87.3	
Bipolaris++	-	-	-	-	-	-	-	-	-	
Chaetomium	-	-	-	-	-	-	-	-	-	
Cladosporium	52	2200	26.3	-	-	-	1	40	3.2	
Curvularia	-	-	-	-	-	-	-	-	-	
Epicoccum	-	-	-	-	-	-	-	-	-	
Fusarium	-	-	-	-	-	-	-	-	-	
Ganoderma	-	-	-	-	-	-	-	-	-	
Myxomycetes++	-	-	-	-	-	-	-	-	-	
Pithomyces++	-	-	-	-	-	-	-	-	-	
Rust	-	-	-	-	-	-	-	-	-	
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-	
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-	
Unidentifiable Spores	-	-	-	-	-	-	-	-	-	
Zygomycetes	-	-	-	-	-	-	-	-	-	
<b>Total Fungi</b>	<b>196</b>	<b>8370</b>	<b>100</b>	-	<b>No Trace</b>	-	<b>30</b>	<b>1260</b>	<b>100</b>	
Hyphal Fragment	-	-	-	-	-	-	-	-	-	
Insect Fragment	-	-	-	-	-	-	-	-	-	
Pollen	-	-	-	-	-	-	-	-	-	
Analyt. Sensitivity 600x	-	42	-	-	0	-	-	42	-	
Analyt. Sensitivity 300x	-	13*	-	-	0*	-	-	13*	-	
Skin Fragments (1-4)	-	-	-	-	-	-	-	3	-	
Fibrous Particulate (1-4)	-	1	-	-	-	-	-	1	-	
Background (1-5)	-	1	-	-	-	-	-	1	-	

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

*Alan Goldstein*  
 Alan Goldstein, Ph.D., Laboratory Manager  
 or other approved signatory

High levels of background particulate can obscure spores and other particulates leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. \*\*\* Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed. EMSL maintains liability limited to cost of analysis. This report relates only to the samples reported above and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. Interpretation and use of test results are the responsibility of the client. Samples received in good condition unless otherwise noted.

Samples analyzed by EMSL Analytical, Inc. Morrisville, NC AIHA-LAP, LLC--EMLAP Lab 173741

Initial report from: 05/22/2019 07:44:55

For information on the fungi listed in this report, please visit the Resources section at [www.emsl.com](http://www.emsl.com)



# EMSL Analytical, Inc.

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### Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number: Client Sample ID: Volume (L): Sample Location	291905207-0004 19-662-04 75 Media Center "Library"			291905207-0005 19-662-05 75 Room 203			291905207-0006 19-662-06 75 Room 305		
	Spore Types	Raw Count	Count/m <sup>3</sup>	% of Total	Raw Count	Count/m <sup>3</sup>	% of Total	Raw Count	Count/m <sup>3</sup>
Alternaria (Ulocladium)	-	-	-	2	80	5.2	-	-	-
Ascospores	3*	40*	8.5	4	200	12.9	14	590	17.3
Aspergillus/Penicillium	2	80	17	1	40	2.6	-	-	-
Basidiospores	8	300	63.8	11	460	29.7	60	2500	73.3
Bipolaris++	-	-	-	-	-	-	1*	10*	0.3
Chaetomium	-	-	-	-	-	-	-	-	-
Cladosporium	4*	50*	10.6	18	760	49	6	300	8.8
Curvularia	-	-	-	-	-	-	-	-	-
Epicoccum	-	-	-	-	-	-	-	-	-
Fusarium	-	-	-	-	-	-	-	-	-
Ganoderma	-	-	-	-	-	-	-	-	-
Myxomycetes++	-	-	-	1*	10*	0.6	1*	10*	0.3
Pithomyces++	-	-	-	-	-	-	-	-	-
Rust	-	-	-	-	-	-	-	-	-
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	-	-	-	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-
<b>Total Fungi</b>	<b>17</b>	<b>470</b>	<b>100</b>	<b>37</b>	<b>1550</b>	<b>100</b>	<b>82</b>	<b>3410</b>	<b>100</b>
Hyphal Fragment	1*	10*	-	-	-	-	-	-	-
Insect Fragment	-	-	-	-	-	-	-	-	-
Pollen	-	-	-	-	-	-	-	-	-
Analyt. Sensitivity 600x	-	42	-	-	42	-	-	42	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-
Skin Fragments (1-4)	-	3	-	-	3	-	-	3	-
Fibrous Particulate (1-4)	-	1	-	-	1	-	-	1	-
Background (1-5)	-	1	-	-	1	-	-	1	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

*Alan Goldstein*

Alan Goldstein, Ph.D., Laboratory Manager  
or other approved signatory

High levels of background particulate can obscure spores and other particulates leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. \*\*\* Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed. EMSL maintains liability limited to cost of analysis. This report relates only to the samples reported above and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. Interpretation and use of test results are the responsibility of the client. Samples received in good condition unless otherwise noted.

Samples analyzed by EMSL Analytical, Inc. Morrisville, NC AIHA-LAP, LLC--EMLAP Lab 173741

Initial report from: 05/22/2019 07:44:55

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### Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number: Client Sample ID: Volume (L): Sample Location	291905207-0007 19-662-07 75 Room 334			291905207-0008 19-662-08 75 Room 240			291905207-0009 19-662-09 75 Cafeteria			
	Spore Types	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total
Alternaria (Ulocladium)	-	-	-	-	-	-	-	-	-	-
Ascospores	22	930	45.1	44	1900	40.3	9	400	11.9	
Aspergillus/Penicillium	-	-	-	2	80	1.7	4	200	6	
Basidiospores	21	890	43.2	58	2400	50.8	39	1600	47.8	
Bipolaris++	-	-	-	-	-	-	-	-	-	
Chaetomium	-	-	-	-	-	-	-	-	-	
Cladosporium	5	200	9.7	7	300	6.4	27	1100	32.8	
Curvularia	-	-	-	-	-	-	-	-	-	
Epicoccum	-	-	-	-	-	-	1*	10*	0.3	
Fusarium	-	-	-	-	-	-	-	-	-	
Ganoderma	-	-	-	-	-	-	-	-	-	
Myxomycetes++	1	40	1.9	1	40	0.8	1	40	1.2	
Pithomyces++	-	-	-	-	-	-	-	-	-	
Rust	-	-	-	-	-	-	-	-	-	
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-	
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-	
Unidentifiable Spores	-	-	-	-	-	-	-	-	-	
Zygomycetes	-	-	-	-	-	-	-	-	-	
<b>Total Fungi</b>	<b>49</b>	<b>2060</b>	<b>100</b>	<b>112</b>	<b>4720</b>	<b>100</b>	<b>81</b>	<b>3350</b>	<b>100</b>	
Hyphal Fragment	1	40	-	5	200	-	1	40	-	
Insect Fragment	-	-	-	-	-	-	-	-	-	
Pollen	-	-	-	1	40	-	1	40	-	
Analyt. Sensitivity 600x	-	42	-	-	42	-	-	42	-	
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-	
Skin Fragments (1-4)	-	2	-	-	3	-	-	4	-	
Fibrous Particulate (1-4)	-	1	-	-	1	-	-	1	-	
Background (1-5)	-	2	-	-	2	-	-	3	-	

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

*Alan Goldstein*

Alan Goldstein, Ph.D., Laboratory Manager  
or other approved signatory

High levels of background particulate can obscure spores and other particulates leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. \*\*\* Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed. EMSL maintains liability limited to cost of analysis. This report relates only to the samples reported above and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. Interpretation and use of test results are the responsibility of the client. Samples received in good condition unless otherwise noted.

Samples analyzed by EMSL Analytical, Inc. Morrisville, NC AIHA-LAP, LLC--EMLAP Lab 173741

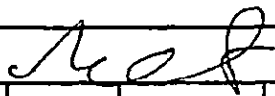
Initial report from: 05/22/2019 07:44:55

For information on the fungi listed in this report, please visit the Resources section at [www.emsl.com](http://www.emsl.com)



Client: ATI Test: M001 Air-O-Cell #Samples: 9  
 Order: 291905207 Project: 19-662-PGCPs - Lake Arbor ES  
 Disposition: Discard after 7/14/2019

ANALYTICAL, INC.  
 130 NORTH  
 ON, NJ 08077  
 (908) 220-3675  
 (908) 786-0262

Company Name: <b>ATI, Inc</b>		EMSL-Bill to: <input type="checkbox"/> Same <input type="checkbox"/> Different if Bill to is Different note instructions in Comments					
Street: 4221 Rumsey Road, Suite 250		Third Party Billing requires written authorization from third party.					
City: Lanham	State/Province: MD	Zip/Postal Code: 20706	Country:				
Report To (Name): Brian Chapman / Mikal Frater		Telephone #: 202-558-7489					
Email Address: Brian@atlin.com & Mikal@atiinc.com		Fax #:	Purchase Order:				
Project Name/Number: 19-662- PGCPs - Lake Arbor ES		Please Provide Results: <input type="checkbox"/> Fax <input checked="" type="checkbox"/> Email					
U.S. State Samples Taken:		Project Zip Code:					
Sterile, Sodium Thiosulfate Preserved Bottle Used: <input type="checkbox"/> Biocide Used in Source (specify): <input type="checkbox"/>		Connecticut Samples: <input checked="" type="checkbox"/> Commercial <input type="checkbox"/> Residential					
Public Water Supply Samples: <input type="checkbox"/> Note: All results may automatically be reported to DOH if required by state.							
Turnaround Time (TAT) Options - Please Check							
<input type="checkbox"/> 3 Hour	<input type="checkbox"/> 6 Hour	<input type="checkbox"/> 24 Hour	<input type="checkbox"/> 48 Hour				
<input type="checkbox"/> 72 Hour	<input type="checkbox"/> 96 Hour	<input checked="" type="checkbox"/> 1 Week	<input type="checkbox"/> 2 Week				
<b>Microbiology Test Codes</b>							
M001 Air-O-Cell	M174 MoldSnap	M012 Pseudomonas aeruginosa (PIA***)	M115 Sewage Screen - Water (PIA***)				
M030 Micro 5	M032 Allergenco-D	M024 Pseudomonas aeruginosa (MFT*)	M116 Sewage Screen - Water (MPN**)				
M041 Fungal Direct Examination		M015 Heterotrophic Plate Count	M117 Sewage Screen - Swab (PIA***)				
M169 Pollen ID & Enumeration		M017 Total Coliform & E. coli (Colilert P/A***)	M013 Sewage Screen - Swab (MFT*)				
M280 Dust Characterization Level-1		M018 Total Coliform & E. coli (MFT*)	M133 Methicillin-resistant Staph. aureus (MRSA)				
M281 Dust Characterization Level-2		M114 Total Coliform & E. coli Enumeration (Colilert MPN**)	M031 Rapid-growing non-TB Mycobacteria Detection & Enumeration				
M005 Viable Fungi- Air Samples (Genus ID & Count)		M019 Fecal Coliform (MFT*)	M014 Endotoxin Analysis				
M006 Viable Fungi- Air Samples (includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)		M020 Fecal Streptococcus (MFT*)	M044 Group Allergen (Cat; Dog, Cockroach, Dust Mite)				
M007 Culturable fungi - Surface Samples (Genus ID & Count)		M029 Enterococci (MFT*)	Other See Analytical Price Guide				
M008 Culturable fungi - Surface Samples (includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)		M129 Enterococci (Enterolert P/A***)	Legionella Analysis Please use EMSL Legionella COC				
M009 Bacteria Culture Gram Stain & Count		M180 Real Time qPCR-ERMI 36 Panel					
M010 Bacteria Count & ID - 3 Most Prominent		M025 Sewage Screen -Water (MFT*)					
M011 Bacteria Count & ID - 5 Most Prominent							
Name of Sampler: Brian Chapman & Mikal Frater		Signature of Sampler: 					
Sample #	Sample Location/Description	Sample Type	Potable/ NonPotable (Only for Waters)	Test Code	Volume/ Area	Date/Time Collected	Temperature (°C) (Lab Use Only)
Example A1	Kitchen Sink/Tap	Water	<input checked="" type="checkbox"/> P <input type="checkbox"/> NP	M017	100 mL	9/1/13 4:00 PM	
19-662-01	Outside Parking Lot	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	75L	05-15-19 - 11:54	
19-662-02	Field Blank	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	75L	05-15-19 -	
19-662-03	Main Office	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	75L	05-15-19 - 12:05	
19-662-04	Media Center "Library"	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	75L	05-15-19 - 12:24	
19-662-05	Room 203	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	75L	05-15-19 - 12:33	
Client Sample # (s): - 9		Total # of Samples: 9		Samples Received Chilled? Yes / No (Lab Use Only)			
Relinquished (Client): <b>MIKAL FRATER</b>		Date: 5-15-19		Time: <b>3:55</b>			
Received (Lab): <b>L. Bennett Walk In</b>		Date: <b>5/15/19</b>		Time: <b>3:55 pm</b>			
Comments/Special Instructions:							

EMSL Analytical, Inc.'s Laboratory Terms and Conditions are incorporated into this chain of custody by reference in their entirety. Submission of samples to EMSL Analytical, Inc. constitutes acceptance and acknowledgment of all terms and conditions by Customer.



**Appendix B:  
Instrument Calibration Records**

# Certificate of Calibration

() Buck™ BioAire Pump Calibration Rotameter

( ) Buck™ BioSlide Pump Calibration Rotameter

Serial number: R14057

Date Calibrated: 1/22/19

Calibration Due Date: 1/22/20

## Flow Calibration

This is to certify that the rotameter listed above has been calibrated using a Buck Primary calibrator listed below which is calibrated according to A.P. Buck, Inc. calibration procedure APB-1, Ver. 6.2 and is traceable to the National Institute of Standards & Technology (N.I.S.T). A.P. Buck guarantees the accuracy of the rotameter to be within  $\pm 5\%$  of the actual flow rate.

AMBIENT CONDITIONS: Temperature  $74 \pm 3^{\circ}$  F Relative Humidity  $50 \pm 10\%$

Description	MFR.	Model	Serial #
Primary Calibrator	A.P. Buck Inc.	M30B	<input type="checkbox"/> A40020 <input checked="" type="checkbox"/> A40021

QA Approval By: 

Information contained in this document should not be reproduced in any form without the written consent of A.P. Buck, Inc. It is for reference only and cannot be used as a form of endorsement by any private or governmental regulatory body.

A.P. BUCK, INC.  
7101 Presidents Drive, Suite 110  
Orlando, FL 32809  
Phone: 407-851-8602  
Fax: 407-851-8910

**BUCK**  
A.P. BUCK, INC.









# CERTIFICATE OF CALIBRATION AND TESTING

TSI Incorporated, 500 Cardigan Road, Shoreview, MN 55126 USA  
 Tel: 1-800-874-2811 1-651-490-2811 Fax: 1-651-490-3824 <http://www.tsi.com>

ENVIRONMENT CONDITIONS				<b>MODEL</b>	<b>982</b>
TEMPERATURE	75.9 (24.4)	°F (°C)		<b>SERIAL NUMBER</b>	<b>P17100006</b>
RELATIVE HUMIDITY	46	%RH			
BAROMETRIC PRESSURE	28.81 (975.6)	inHg (hPa)			

<input checked="" type="checkbox"/> AS LEFT	<input checked="" type="checkbox"/> IN TOLERANCE
<input type="checkbox"/> AS FOUND	<input type="checkbox"/> OUT OF TOLERANCE

- CALIBRATION VERIFICATION RESULTS -

TEMPERATURE VERIFICATION				SYSTEM T-101			Unit: °F (°C)
#	STANDARD	MEASURED	ALLOWABLE RANGE	#	STANDARD	MEASURED	ALLOWABLE RANGE
1	32.0 (0.0)	32.4 (0.2)	31.0-33.0 (-0.6-0.6)	2	140.0 (60.0)	140.8 (60.4)	139.0-141.0 (59.4-60.6)

HUMIDITY VERIFICATION				SYSTEM H-102			Unit: %RH
#	STANDARD	MEASURED	ALLOWABLE RANGE	#	STANDARD	MEASURED	ALLOWABLE RANGE
1	10.0	9.6	7.8-12.2	4	70.0	69.7	67.8-72.2
2	30.0	29.7	27.8-32.2	5	90.0	89.3	87.8-92.2
3	50.0	49.9	47.8-52.2				

CO2 GAS VERIFICATION				SYSTEM G-101			Unit: ppm
#	STANDARD	MEASURED	ALLOWABLE RANGE	#	STANDARD	MEASURED	ALLOWABLE RANGE
1	0	0	0-50	4	3031	3043	2940-3122
2	518	510	468-568	5	5000	4988	4850-5150
3	1020	1030	970-1070				

CO GAS VERIFICATION				SYSTEM G-101			Unit: ppm
#	STANDARD	MEASURED	ALLOWABLE RANGE	#	STANDARD	MEASURED	ALLOWABLE RANGE
1	36	36	33-39	2	101	100	98-104

*TSI does hereby certify that the above described instrument conforms to the original manufacturer's specification (not applicable to As Found data) and has been calibrated using standards whose accuracies are traceable to the United States National Institute of Standards and Technology (NIST) or has been verified with respect to instrumentation whose accuracy is traceable to NIST, or is derived from accepted values of physical constants. TSI's calibration system is registered to ISO-9001:2015.*

Measurement Variable	System ID	Last Cal.	Cal. Due	Measurement Variable	System ID	Last Cal.	Cal. Due
Temperature	E003986	02-14-18	08-31-18	Temperature	E003987	02-14-18	08-31-18
Humidity	E003539	02-22-18	08-31-18	5000 CO2	c5732043	04-16-18	10-04-20
200 CO	CC506122	01-24-18	01-25-26	N2	t78516	04-17-18	04-03-23
Air	108551y	04-23-18	03-09-20	Flow	E003298	10-25-17	10-31-18
Flow	E004631	10-25-17	10-31-18	Flow	E003980	03-28-18	03-31-19
Flow	E003525	01-10-18	01-31-19	2000 C4H8	EB0053919	10-20-17	10-20-21
100 C4H8	EB0078607	09-28-16	09-28-20				

Chimona

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CALIBRATED

May 29, 2018

DATE